University of Delaware Biosafety Committee
Meeting Minutes- April 1, 2026 11:30am-1:30pm
Hybrid- In Person and Zoom Meeting

Present	Member
	Dr. Jennifer Biddle, School of Marine Science and Public Policy
X	Dr. Erin Brannick, Animal and Food Sciences, Animal Expert, IBC Chair
X	Ms. Renee Brown, Research Office
X	Dr. Brandon Calitree, Environmental Health and Safety
	Dr. Nicole Donofrio, Plant and Soil Sciences, Plant Expert
X	Ms. Michelle Ferguson, Biosafety Officer, Environmental Health and Safety
X	Ms. Michelle Hamilton, Community Member
	Mr. Norm Henry, Community Member
	Dr. Brian Kwee, Biomedical Engineering
	Dr. Anja Nohe, Biological Sciences
	Dr. Mark Parcells, Animal and Food Sciences, Animal Expert
X	Ms. Margaret (Meg) Roth, School of Nursing
	Dr. Stephen Streatfield, Plant and Soil Sciences, Plant Expert
X	Ms. Heather Walters, Medical and Molecular Sciences
X	Dr. Neal Zondlo, Chemistry and Biochemistry

A quorum was met for this meeting.

Guests Present
Ms. Diana LaPier, Environmental Health and Safety

Call to Order

- Dr. Brannick called the meeting to order at 11:30am. Minutes from January 15, 2026 meeting were approved. (8 for/0 against)

Committee Review of Recombinant DNA Registrations

- The Committee reviewed the list of exempt category research. Fourteen protocols met this category (26-009, 26-010, 26-011, 26-012, 26-013, 26-014, 26-016, 26-017, 26-018, 26-020, 26-022, 26-023, 26-024, 26-025). The Committee voted unanimously in favor of acceptance.

- The non-exempt research was reviewed next. Three experiments met this category. 26-015, 26-019, and 26-021 were reviewed as indicated on the Review Forms beginning on page 3. The protocols were unanimously voted in favor of acceptance.

Incident Review

- The committee reviewed four incidents, I26-001, I26-002, I26-003, I26-004 that were deemed non-reportable according to the NIH Guidelines

Program Summaries and Ongoing Oversight

The committee discussed updates within the select agent program, bloodborne pathogen program and biosafety program.

- The University had its renewal inspection of the select agent program in January 13-14. The committee discussed the findings and the registration renewal for the program.
- Discussion of registration compliance for the BBP and Biosafety programs, as well as training deficiencies were presented to the committee. Individuals have been contacted monthly regarding their outstanding items for each program.

Old Business

- UBC SOP revisions and updates were discussed regarding adding in information about NIH transparency for IBC. Since the NIH transparency is still ongoing, the Committee deferred finalization until the June 2026 meeting.

New Business

- The BSO notified the committee that the rDNA webform has been updated to include a general description field that will be used for the meeting minutes.
- The committee also discussed reviewing and updating the injury form for EHS as well as the flow charts for injuries. A draft of the flow chart will be reviewed at the next meeting in June.

IBC Training

- The Committee discussed escalation for delinquent training.

Public Comments

- There were no public comments for this meeting.

Adjournment

- The Committee moved to adjourn the meeting, unanimously approved at 12:24 p.m.
- The next meeting will be held on Thursday June 4, at 11:30

Respectfully submitted,
Michelle Ferguson
Michelle Ferguson
Biosafety Officer

University Biosafety Committee Review Form
For Recombinant DNA Experiments Covered by the
NIH *Guidelines*

Principal Investigator: _____Dr. Arit Ghosh_____

Department: Bioimaging Center

Project Title: Cell Line Construction via TetOn Lentiviral Plasmid

Registration Number: _____26-015 (3 year renewal 23-048)

NIH Guidelines Section: III-E-1

Description of procedures provided: Yes __XX__ No _____
Tracking ER-phagy mediated Tcell killing of tumor cells via flow cytometry requires innovative approaches. In the core faciltiy, we are developing a novel assay which employs an mCherry-eGFP-RAMP4 construct delivered into MCF7 cancer cell lines. It acts as a stable membrane protein to visualize and quantify ER-phagy, which we hypothesize can be degraded via the lysosome, if the host cancer cells are targeted via lysosomal degrdation as evidenced in previous studies. The shifts in tandem fluorescence, termed as EATR assays, will be quantified as ER turnover in an inducible Tet-On-system in turn affected by T cell mediated killing in co-cultures.

Containment Level: _____BSL-2_____

Appropriate facilities to be used: Yes __XX___ No _____

Procedures acceptable for containment: Yes ___XX____ No _____

Work practices acceptable for containment: Yes __XX_____ No _____

Training/ Experience of Personnel acceptable for work: Yes ___XX____ No ____

Comments: The committee agreed with the proposed procedures, work practices and containment. Lab members listed on this protocol have been properly trained with techniques. No concerns were brought up.

These items have been reviewed by the University Biosafety Committee and the committee has voted **__XX_ For** _____ Against approval of this project on this date. (10 for/0 against)
Experiments covered by this protocol can now be initiated.

UBC Representative: _____*Michelle L Ferguson*_____
Date: _____4/1/26_____

University Biosafety Committee Review Form
For Recombinant DNA Experiments Covered by the
NIH *Guidelines*

Principal Investigator: _____Dr. Austin Keeler_____

Department: _Biological Sciences

Project Title: i-GONAD gene editing of small mammals in the DCMBS COBRE Transgenic Rodent Core (TGRC)

Registration Number: _____26-019 (new protocol)____

NIH Guidelines Section: III-E-4

Description of procedures provided: Yes __XX__ No _____
The DCMBS COBRE Transgenic Rodent Core (TGRC) is intended to be a small, agile core to generate gene edited small mammals for biomedical research. This animal protocol is intended to be a standard protocol to allow the core to generate gene edited research mammals as needed to support research without causing undue burden on the core, research labs and PIs, OLAM, and IACUC with extensive protocol revisions and deletions to add the i-GONAD when generating new lines, while ensuring animals do not multiple procedures. i-GONAD, or improved Genome-editing via Oviductal Nucleic Acid Delivery, is a new method to produce transgenic mammals that performs gene editing *in vivo*, in the oviduct of recently impregnated females, bypassing the expensive equipment and highly skilled personnel required in conventional transgenic cores. No specific models or edits are proposed here, simply the workflow and procedure, to allow the UD IACUC to advise on how to coordinate use of the TGRC within the current IACUC system.

Containment Level: _____BSL-2_____

Appropriate facilities to be used: Yes __XX___ No _____

Procedures acceptable for containment: Yes ___XX____ No _____

Work practices acceptable for containment: Yes __XX_____ No _____

Training/ Experience of Personnel acceptable for work: Yes ___XX____ No ____

Comments: The committee agreed with the proposed procedures, work practices and containment. Lab members listed on this protocol have been properly trained with techniques. No concerns were brought up.

These items have been reviewed by the University Biosafety Committee and the committee has voted **__XX_ For** _____ Against approval of this project on this date. (13 for/0 against)

Experiments covered by this protocol can now be initiated.

UBC Representative: _____*Michelle L Ferguson*_____
Date: _____4/1/26_____

University Biosafety Committee Review Form
For Recombinant DNA Experiments Covered by the
NIH *Guidelines*

Principal Investigator: _____Dr. Mark Parcells_____

Department: Animal and Food Sciences

Project Title: Use of Lentiviral Vectors to Assess Marek's Disease Virus and Chicken Cellular Gene Function

Registration Number: _____26-021 (3 year renewal of 23-009)____

NIH Guidelines Section: III-E-1

Description of procedures provided: Yes __XX__ No _____
The purpose of this work is the construction of stable expression cell lines for projects involving Marek's disease virus (MDV) and its host cell interactions. We are focusing on the interaction of the viral oncoprotein Meq, its isoforms and splice variants, and the cellular proteins with which they interact. In the next few years, we are planning to examine Meq proteins from high virulence strains (very virulent and very virulent plus) and their interactions with cellular DNA repair and damage-associated proteins.

Containment Level: _____BSL-2_____

Appropriate facilities to be used: Yes __XX___ No _____

Procedures acceptable for containment: Yes ___XX____ No _____

Work practices acceptable for containment: Yes __XX_____ No _____

Training/ Experience of Personnel acceptable for work: Yes ___XX____ No ____

Comments: The committee agreed with the proposed procedures, work practices and containment. Lab members listed on this protocol have been properly trained with techniques. No concerns were brought up.

These items have been reviewed by the University Biosafety Committee and the committee has voted **__XX_ For** _____ Against approval of this project on this date. (13 for/0 against)
Experiments covered by this protocol can now be initiated.

UBC Representative: _____*Michelle L Ferguson*_____

Date: _____4/1/26_____